Enzyme Immunoassay for
Corticosterone

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INTRODUCTION

Corticosterone is a glucocorticoid secreted by the adrenal cortex in response to stress. Glucocorticoids are also essential for proper metabolism of fats, proteins and carbohydrates in the body. While cortisol and corticosterone are both produced in response to stress in humans, corticosterone is the predominant glucocorticoid produced in mice and rats. Corticosterone has also been used as a predictor of stress in a variety of wild animals. In particular, corticosterone levels were increased in Galapagos marine iguanas under famine conditions brought on by El Nino. While not always predictive of environmental stress in the wild[2], conditions such as starvation, rapid temperature change or changes in daily routine can cause increased corticosterone levels in laboratory mice and rats.

Corticosterone is also administered to treat inflammation because like many other steroids, it is a powerful anti-inflammatory agent.

PRINCIPLES OF PROCEDURE

This is an ELISA (Enzyme-Linked Immunosorbent Assay) for the quantitative analysis of Corticosterone levels in biological fluid. This test kit operates on the basis of competition between the enzyme conjugate and the corticosterone in the sample for a limited number of binding sites on the antibody coated plate.

The sample or standard solution is first added to the microplate. Next, the diluted enzyme conjugate is added and the mixture is shaken and incubated at room temperature for one hour. During the incubation, competition for binding sites is taking place. The plate is then washed, removing all the unbound material. The bound enzyme conjugate is detected by the addition of the substrate that generates an optimal color after 30 minutes. Quantitative test results may be obtained by measuring and comparing the absorbance reading of the sample wells against the wells of the standard curve with a microplate reader at 650 nm. The extent of color development is inversely proportional to the amount of corticosterone in the sample or standard. For example, the absence of corticosterone in the sample will result in a bright blue color, whereas the presence of corticosterone will result in decreased or no color development.

MATERIALS PROVIDED

Component	Description	Volume	Storage
EIA Buffer	Buffer used to dilute the Conjugate and Corticosterone Standards.	30 mL	4°C
10x Wash Buffer	Buffer used to wash the plate prior to color development.	20 mL	4°C
TMB Substrate	TMB substrate used for color development.	20 mL	4°C
5x Extraction Buffer	Buffer used to dilute extracted and non-extracted samples.	30 mL	4°C
Corticosterone-HRP Conjugate	Corticosterone horseradish peroxidase concentrated conjugate.	150 µL	4°C
Corticosterone Standard	1 µg/mL Corticosterone standard solution in methanol.	100 µL	4°C
Coated Plate	96-well microplate coated with rabbit anti-Corticosterone antibody.	1 plate	4°C

MATERIALS NEEDED BUT NOT PROVIDED

1. Microplate reader with a 450 nm or 650 nm filter
2. Adjustable micropipettes (10 – 1000 μL) and tips
3. Deionized water
4. Plate cover or plastic film
5. Test tubes
6. 1 N HCl (optional)

EXTRACTION MATERIALS

1. Ethyl Ether
2. Nitrogen Gas
3. Vortex

STORAGE

1. Store the components of this kit at the temperatures specified on the labels.
2. Unopened reagents are stable until the indicated kit expiration date.
3. Desiccant bag must remain in foil pouch with unused strips. Keep pouch sealed when not in use to maintain a dry environment. Remove excess air before sealing.

WARNINGS AND PRECAUTIONS

1. Use aseptic technique when opening and dispensing reagents.
2. This kit is designed to work properly as provided and instructed. Additions, deletions or substitutions to the procedure or reagents are not recommended, as they may be detrimental to the assay.

PROCEDURAL NOTES

1. The enzyme conjugate is most stable in its concentrated form. Dilute only the volume necessary for the amount of strips currently being used.
2. To minimize errors in absorbance measurements due to handling, wipe the exterior bottom of the microplate wells with a lint-free paper towel prior to inserting into the plate reader.

SAMPLE PREPARATION

1. Urine and tissue culture supernatant can be assayed after diluting them with diluted Extraction Buffer.
2. Plasma and most other mediums will need to be extracted.

EXTRACTION PROTOCOL

1. Pipet 100 µL of plasma into a glass test tube (10x75 mm) and add 1 mL of Ethyl Ether.
2. Vortex the tube for 30 seconds and allow the phases to separate.
3. Transfer the organic phase into a clean glass tube and evaporate the solvent with a Nitrogen stream.
4. Dissolve the residue in 100 µL of diluted Extraction Buffer.
5. Dilute the extract 100-fold by adding 10 µL of the above extract into 990 µL of diluted Extraction Buffer.
6. If the concentration is higher than the high range of the standard curve, the samples will need to be further diluted and re-assayed.

REAGENT PREPARATION

1. **5x Extraction Buffer:** Dilute the appropriate amount to 1x with deionized water prior to use.
2. **10x Wash Buffer:** Add 20 mL of 10x Wash Buffer to 180 mL of deionized water prior to use.
3. **Corticosterone-HRP Conjugate:** Dilute 110 µL of Conjugate into 5.5 mL total volume of EIA Buffer.

STANDARD CURVE PREPARATION

The Corticosterone Standard is provided as a 1 µg/mL stock solution in methanol. Use the following tables to dilute a set of standard stock solutions and construct an eight-point standard curve.

Table 1: Standard Stock Preparation

Standard	Corticosterone Conc. (ng/mL)	Vol. of EIA Buffer (µL)	Transfer Vol. (µL)	Final Vol. (µL)
A	1000	-	Provided	80
B	20	980	20 µL of A	800
C	2	1800	200 µL of B	1800
D	0.2	1800	200 µL of C	2000

Table 2: Standard Curve Preparation

Standard	Corticosterone Conc. (ng/mL)	Vol. of EIA Buffer (µL)	Vol. of Stock B (µL)	Vol. of Stock C (µL)	Vol. of Stock D (µL)
S_0	0	1000	-	-	-
S_1	0.05	750	-	-	250
S_2	0.1	500	-	-	500
S_3	0.2	-	-	-	1000
S_4	0.5	750	-	250	-
S_5	1.0	500	-	500	-
S_6	2.0	-	-	1000	-
S_7	5.0	750	250	-	-

ASSAY PROCEDURE

1. Add 50 µL of Standards or Samples to the corresponding wells on the microplate in duplicate. See **Scheme I** for a sample plate layout.
2. Add 50 µL of diluted Corticosterone-HRP Conjugate to each well. Incubate at room temperature for one hour.
3. Wash the plate three times with 300 µL of diluted Wash Buffer per well. Wash 5 times if using an automated plate washer.
4. Add 150 µL of TMB Substrate to each well. Incubate at room temperature for 30 minutes.
5. Read the plate at 650 nm.
   Alternately, the color reaction can be stopped after 10-15 minutes by adding 50 µL of 1 N HCl and read at 450 nm.
   NOTE: If accounting for substrate background, use 2 wells as blanks (BLK) with only 150 µL TMB Substrate in the wells. Subtract the average of these absorbance values from the absorbance values of the wells being assayed.

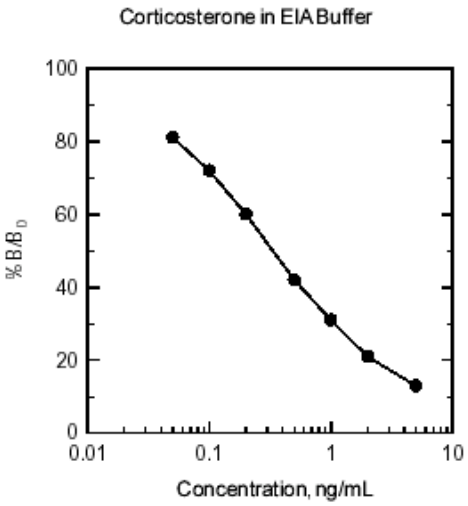
Scheme I: Sample Plate Layout

	1	2	3	4	5	6	7	8	9	10	11	12
A	S_0	S_0	U_1	U_1	U_9	U_9	U_{17}	U_{17}	U_{25}	U_{25}	U_{33}	U_{33}
B	S_1	S_1	U_2	U_2	U_{10}	U_{10}	U_{18}	U_{18}	U_{26}	U_{26}	U_{34}	U_{34}
C	S_2	S_2	U_3	U_3	U_{11}	U_{11}	U_{19}	U_{19}	U_{27}	U_{27}	U_{35}	U_{35}
D	S_3	S_3	U_4	U_4	U_{12}	U_{12}	U_{20}	U_{20}	U_{28}	U_{28}	U_{36}	U_{36}
E	S_4	S_4	U_5	U_5	U_{13}	U_{13}	U_{21}	U_{21}	U_{29}	U_{29}	U_{37}	U_{37}
F	S_5	S_5	U_6	U_6	U_{14}	U_{14}	U_{22}	U_{22}	U_{30}	U_{30}	U_{38}	U_{38}
G	S_6	S_6	U_7	U_7	U_{15}	U_{15}	U_{23}	U_{23}	U_{31}	U_{31}	U_{39}	U_{39}
H	S_7	S_7	U_8	U_8	U_{16}	U_{16}	U_{24}	U_{24}	U_{32}	U_{32}	BLK	BLK

CALCULATIONS

1. After the substrate background has been subtracted from all absorbance values, average all of your duplicate well absorbance values.
2. The average of your two S_0 values is now your B_0 value. (S_1 now becomes B_1, etc.)
3. Next, find the percent of maximal binding (%B/B_0 value). To do this, divide the averages of each standard absorbance value (now known as B_1 through B_7) by the B_0 absorbance value and multiply by 100 to achieve percentages.
4. Graph your standard curve by plotting the %B/B_0 for each standard concentration on the y-axis against concentration on the x-axis. Draw a curve by using a curve-fitting routine (i.e. 4-parameter or linear regression).
5. Divide the averages of each sample absorbance value by the B_0 value and multiply by 100 to achieve percentages.
6. Using the standard curve, the concentration of each sample can be determined by comparing the %B/B_0 of each sample to the corresponding concentration of corticosterone standard.
7. If the samples were diluted, the concentration determined from the standard curve must be multiplied by the dilution factor.

Figure 1: Typical Standard Curve



CROSS REACTIVITY

Corticosterone	100.0%	21-Desoxycortisol	0.24%
Deoxycorticosterone	38.0%	d-Aldosterone	0.13%
6-Hydroxycorticosterone	19.0%	Testosterone	0.12%
Progesterone	5.1%	17α-Hydroxyprogesterone	0.12%
Tetrahydrocorticosterone	2.7%	Prednisone	0.10%
Prednisolone	1.5%	Dexamethasone	0.03%
Cortisol	1.1%	Cholesterol	<0.01%
Pregnenolone	0.85%	Estradiol	<0.01%
11-Epicorticosterone	0.78%	Estriol	<0.01%
Cortisone	0.27%		

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